

Avian Colibacillosis - A Mini Review

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ABSTRACT

Avian colibacillosis is an economically important disease of poultry throughout the world. It causes great economic losses to the poultry farmers throughout the world in terms of mortality of affected birds, decreased productivity, increased costs towards treatment of affected birds etc. This article briefly review about epidemiology, host range, clinical signs, diagnosis of avian colibacillosis.

Key words: Avian colibacillosis, poultry, *E. coli*

INTRODUCTION

Avian colibacillosis refers to any localized or systemic infection caused entirely or partly by avian pathogenic *Escherichia coli* (APEC), including colisepticaemia, coligranuloma (Hjarre's disease), air sac disease, swollen-head syndrome, venereal colibacillosis and coliform cellulitis, peritonitis, salphingitis, orchitis, osteomyelitis/synovitis, panophthalmitis, omphalitis/yolk sac infection and enteritis³.

Avian colibacillosis is an infectious disease of birds caused by *Escherichia coli* (*E. coli*) which is considered as one of the principal causes of morbidity and mortality, associated with heavy economic losses to the poultry industry by its association with various disease conditions, either as primary pathogen or as a secondary pathogen¹¹.

Occurrence

Global scenario

Colibacillosis in chicken was reported from various parts of the world including Bangladesh¹⁸, Belgium²⁶, China⁷, Denmark¹³, Japan²², Korea¹², and United States of America²⁹.

Indian scenario

Escherichia coli was isolated from tissues of chicken and antibiotic resistance pattern was studied by Sharada and Wilfred Ruban²⁰, at Karnataka. Detection and characterisation of shiga toxin producing *E. coli* (STEC) and enteropathogenic *E. coli* (EPEC) in poultry was carried out by Dutta *et al*⁶.

Pathogenicity studies of twenty one *E. coli* isolates recovered from clinical cases of colibacillosis in chickens at Uttar Pradesh revealed that seventeen of them were found pathogenic²⁵. Sarma *et al*¹⁹, reported that ninety nine strains of *E. coli* isolates from domestic fowl were serotyped into 56 different serotypes in Punjab.

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Escherichia coli associated egg peritonitis was responsible for 15.39 per cent of the reproductive tract abnormalities in commercial layers between 21 and 80 weeks of age in Namakkal area of Tamil Nadu for the period from 2005 to 2008. Drop in egg production and mortality in these layer flocks varied from 3 to 20 per cent and 0.5 to 7 per cent²³, respectively.

Epidemiology

Agent

Escherichia genus consists of gram negative, non-acid-fast, uniform staining, non-spore forming bacillus, usually of 2-3 X 0.6 µm in size belonging to the family Enterobacteriaceae³. It is considered as a member of normal microflora of the poultry intestine, but certain strains such as avian pathogenic *E. coli* (APEC), spread to various internal organs and cause colibacillosis characterised by systemic fatal disease¹⁰. Serotypes of *E. coli* are classified according to the Kauffmann scheme. At present there are approximately 180 O, 60 H, and 80 K antigens were recognized²⁴. In most serologic typing schemes only the O and H antigens are determined³.

In domestic poultry, avian colibacillosis is frequently associated with *E. coli* strains of serotypes O78:K80, O1: K1 and O2: K1¹¹.

The occurrence of a specific serotype and its role in disease production depends upon the health status of the birds, climatic conditions, geographical situations and managerial strategies²³.

Host

Most, if not all, avian species are susceptible to colibacillosis. Clinical disease is reported most often in chickens, turkeys and ducks. Collectively, the various forms of colibacillosis are considered to be the most common infectious bacterial disease of broiler chickens and turkeys³. Avian pathogenic *E. coli* infection is most frequent in breeders followed by broilers and layers respectively and it is responsible for both embryo as well as early chick mortality in breeders⁹. Colisepticaemia is the commonest infectious

disease of farmed poultry seen worldwide in chicken, turkeys etc⁴.

Risk factors associated with occurrence of avian colibacillosis

Age

All ages are susceptible to colibacillosis, but young birds are more frequently affected and severity of the disease is greater in young birds including developing embryos⁸.

Avian colibacillosis is prevalent in all age groups of chickens (9.52 to 36.73 %) with especially high prevalence rate in adult layer birds (36.73 %)¹⁷. Most outbreaks of colibacillosis occur around the period of peak production²⁶.

Avian pathogenic *E. coli* are responsible for a considerable number of various diseases at different ages including neonatal infection of chicks, oophoritis or salphingitis in laying hen, air sacculitis and septicaemia in all ages of chickens¹¹.

The incidence of the egg peritonitis caused by *E. coli* was noticed throughout the laying period, however more common during the peak production *i-e.*, 21-60 weeks of age²³.

Host susceptibility factors

When hens are beak trimmed at a younger age, their beaks may regrow, and pecking can lead to cannibalism results in *E. coli* infection².

High egg production is associated with an imbalance between the oestrogen and the progesterone levels, which causes reduced resistance to ascending infections on the oviduct²⁶.

Various intrinsic and extrinsic factors like damage to mucosal and skin barriers, impaired mononuclear phagocytic system, immuno-suppression, extreme environmental temperature, stress and even the effects of vaccine viruses like Newcastle disease virus (NDV), infectious bronchitis virus (IBV) and infectious bursal disease (IBD) have been correlated with pathogenicity of *E. coli* in chickens²⁵. Infectious laryngotracheitis virus, Marek's disease virus, influenza virus, chicken infectious anaemia virus, *Ascardia galli*, *Eimeria* species predisposes birds to colibacillosis³. Several pathogens like NDV,

IBV and *Mycoplasma gallisepticum* may play a part in the occurrence of colibacillosis¹¹.

Clinical signs

Pathogenic *E. coli* infection in birds may cause colisepticaemia, air sac disease, pericarditis and perihepatitis, mushy chick disease, acute septicaemia etc²¹. Clinical signs seen in birds affected with colisepticaemia are diarrhea, pasty vent, loss of appetite, depression, dyspnea and sneezing²⁷. In air sac disease affected birds have cloudy, thickened, oedematous air sacs with caseous deposition¹. Clinical signs vary from inapparent to total unresponsiveness just prior to death depending on the specific type of disease produced by *E. coli*. Localized infections generally result in fewer and milder clinical signs than systemic diseases. Affected birds are typically undersized for the flock and found at the ends of the house along the side walls, or under feeder or waterers. Young birds with omphalitis and infected yolk sacs also may have difficulty in walking. Birds with colisepticaemia are often terminally moribund and the flock may be inactive and anorectic. They sit with their eyes closed in a hunched position with drooping of the head, neck and wings. Although, technically, death is not a clinical sign, this may be the main indication of an outbreak of colibacillosis in a flock. Clinical signs of predisposing or compounding factors often are seen concurrently with signs of *E. coli* infections³.

Gross lesions

Gross lesions characteristic of colisepticaemia in birds inclusive of fibrinous perihepatitis, pericarditis, congestion and regression of ovarian follicles, mild to moderate flaccid and wrinkled ovarian follicles with rupture of theca wall resulting in egg peritonitis, salpingitis¹². Rupture of ovarian follicles, presence of amorphous yolk material in the peritoneal cavity, more commonly attached to the surface of the ova and the serous surface of the intestines and oviduct through strands were the gross lesions observed in egg peritonitis associated with *E. coli* infection²³.

Diagnosis

Collection of samples

The diagnosis of colibacillosis is by isolation of *E. coli* from cardiac blood and affected tissues like liver, spleen, pericardium or bone marrow¹¹. Bone marrow cultures are easy to obtain and are generally free of contaminating bacteria³. Swabs collected from liver, oviduct and body cavity were also used for isolation of *E. coli* from colibacillosis affected birds¹².

Isolation and identification

Selective media like MacConkey, eosin-methylene blue (EMB) or drigalki agar are used for isolation of *E. coli*¹¹. A presumptive diagnosis of *E. coli* infection can be made if most of the colonies are characteristically dark with metallic sheen on EMB agar, bright pink with precipitate surrounding colonies on MacConkey agar, or yellow on tergitol-6 agar³. Further identification of the isolated colonies is based on biochemical reactions (indole production, fermentation of glucose with gas production, presence of beta-galactosidase, absence of hydrogen sulphide production and urease, and the inability to utilize citrate as carbon source⁵. *Escherichia coli* was identified as gram negative rods by Gram staining technique¹².

Economic losses

Colibacillosis is one of the main causes of economic loss in the poultry industry worldwide². Despite being known for over a century, avian colibacillosis remains one of the major endemic diseases of poultry resulting in decrease in productivity, mortality and economic losses¹⁵. About 5.5 per cent mortality and 10-20 per cent drop in eggs was observed with *E. coli* infections in egg type layers reared in cages¹⁶. Heavy economic loss occurs in colibacillosis affected broilers and layers due to morbidity and mortality, reduced production and poor chick quality¹⁷. Omer *et al.*¹⁴, reported that colibacillosis in layers caused 1.9 per cent mortality, results in 824.6 US \$ economic loss in layers.

REFERENCES

1. Al Ankari, A.R., J.M. Bradbury, C.J. Naylor, K.J. Worthington, C. Payne Johnson and R.C. Jones, Avian pneumovirus infection in broiler chicks

- inoculated with *Escherichia coli* at different time intervals. *Avian Pathol.*, **30**: 257-267 (2001).
2. Barnes, H.J. and W.B. Gross, Colibacillosis. In: B.W. Calnek (ed.), Diseases of poultry, 10th edn., Iowa State University Press, Ames, USA. pp. 131-141 (1997).
 3. Barnes, J.H., L.K. Nolan and J.P. Vaillancourt, Colibacillosis. In: Saif, Y.M., A.M. Fadly, J.R. Glisson, L.R. McDougald, L.K. Nolan and D.E. Swayne, (ed.), Diseases of poultry, 12th edn., Blackwell publishing professional, Ames, Iowa, USA. pp: 691-737 (2008).
 4. Dhama, K., S. Chakraborty, R. Barathidasan, R. Tiwari, S. Rajagunalan and S. Dayal Singh, *Escherichia coli*, an economically important avian pathogen, its disease manifestations, diagnosis and control, and public health significance: A review. *Res. Opin. Anim. Vet. Sci.*, **3**: 179-194 (2013).
 5. Dho-Moulin, M. and J.M. Fairbrother, Avian pathogenic *Escherichia coli* (APEC). *Vet. Res.*, **30**: 299-316 (1999).
 6. Dutta, T.K., P. Roychoudhury, S. Bandyopadhyay, S.A. Wani and I. Hussain, Detection and characterization of shiga toxin producing *E. coli* (STEC) and enteropathogenic *E. coli* (EPEC) in poultry birds with diarrhea. *Indian J. Med. Res.*, **133**: 541-545 (2011).
 7. Jiang, H.X., D.H. Lu, Z.L. Chen, X.M. Wang, J.R. Chen, Y.H. Liu, X.P. Liao, J.H. Liu and Z.L. Zeng, High prevalence and widespread distribution of multi-resistant *Escherichia coli* isolates in pigs and poultry in China. *Vet J.*, **187**: 99-103 (2011).
 8. Johnson, L.C., S.F. Bilgili, F.J. Hoerr, B.L. McMurtrey and R.A. Norton, The influence of *Escherichia coli* strains from different sources and age of broiler chickens on the development of cellulitis. *Avian Pathol.*, **2**: 38-49 (2001).
 9. Khoo, L.L., Y. Hasnath, Y. Rosnah, N. Saiful, M.A. Maswathi, and M. Ramlan, The prevalence of avian pathogenic *Escherichia coli* (APEC) in peninsular Malaysia. *Malaysian J. Vet. Res.*, **1**: 27-31 (2010).
 10. La Ragione, R.M. and M.J. Woodward, Virulence factors of *Escherichia coli* serotypes associated with avian colisepticaemia. *Res. Vet. Sci.*, **73**: 27-35 (2002).
 11. Lutful Kabir, S.M., Avian colibacillosis and Salmonellosis: A closer look at epidemiology, pathogenesis, diagnosis, control and public health concerns. *Int. J. Environ. Res. Public Health*, **7**: 89-114 (2010).
 12. Oh, J.Y., M.S. Kang, J.M. Kim, B.K. An, E.A. Song, J.Y. Kim, E.G. Shin, M.J. Kim, J.H. Kwon and Y.K. Kwon, Characterization of *Escherichia coli* isolates from laying hens with colibacillosis on two commercial egg-producing farms in Korea. *Poult. Sci.*, **90**: 1948-1954 (2011).
 13. Olsen, R.H., N.M. Stockholm, A. Permin, J.P. Christensen, H. Christensen and M. Bisgaard, Multi-locus sequence typing and plasmid profile characterization of avian pathogenic *Escherichia coli* associated with increased mortality in free range layer flocks. *Avian Pathol.*, **40**: 437-444 (2011).
 14. Omer, M.M., S.M. Adusalab, M.M. Gumaa, S.A. Mulla, E.A. Omer, I.E. Jeddah, A.M. Al-Hassan, M.A. Hussein and A.M. Ahmed, Outbreak of colibacillosis among broiler and layer flocks in intensive and semi intensive poultry farms in Kassala state, Eastern Sudan. *Asian J. Poult. Sci.*, **4**: 173-181 (2010).
 15. Otaki, Y., Poultry disease control programme in Japan. *Asian Livestock*, **20**: 65-67 (1995).
 16. Qu, F.Q., S.Q. Yang, B. Liu, D.Z. Zhang, H.W. Li, S. Liu and S.Y. Chen, Diagnosis and control of swollen head syndrome in layers. *Chinese J. Vet. Med.*, **23**: 23 (1997).
 17. Rahman, M.A., M.A. Samad, M.B. Rahman and S.M.L. Kabir., Bacterio-

- pathological studies on salmonellosis, colibacillosis and pasteurellosis in natural and experimental infections in chickens. *Bangladesh J. Vet. Med.*, **2**: 1-8 (2004).
18. Rashid, M.H., Xue, C., M.R. Islam, M.T. Islam and Y. Cao, A longitudinal study on the incidence of mortality of infectious diseases of commercial layer birds in Bangladesh. *Prev. Vet. Med.*, **109**: 354-358 (2013).
19. Sarma, D.K., D.S. Sambyal and S.N. Sharma, *Escherichia coli* serotypes in domestic fowl of Punjab. *British Vet. J.*, **143**: 273-277 (1987).
20. Sharada, R. and S. Wilfred Ruban, Isolation, characterization and antibiotic resistance pattern of *E. coli* isolated from poultry. *American-Eurasian J. Sci. Res.*, **5**: 18-22 (2010).
21. Singh, S.D., R. Tiwari and K. Dhama, Avian colibacillosis, an economically important disease of young chicks. *Poultry world*, October issue, pp.14-20 (2011).
22. Someya, A., K. Otsuki and T. Murase, Characterization of *Escherichia coli* strains obtained from layer chickens affected colibacillosis in a commercial egg producing farm. *J. Vet. Med. Sci.*, **69**: 1009-1014 (2007).
23. Srinivasan, P., G.A. Balasubramaniam, T.R. Gopalakrishna murthy and P. Balachandran, Bacteriological and pathological studies of egg peritonitis in commercial layer chicken in Namakkal area. *Asian Pacific J. Trop. Biomed.*, **3**: 988-994 (2013).
24. Stenutz, R., A. Weintraub and G. Widmalm, The structures of *Escherichia coli* O-polysaccharide antigens. *FEMS. Microbiol. Rev.*, **30**: 382-403 (2006).
25. Sylvester, S.A., S.D. Singh and M. Mahender, *In-Ovo* and *In-Vivo* pathogenicity study of avian *Escherichia coli* isolated from cases of colibacillosis in chickens. *Indian J. Vet. Pathol.*, **32**: 43-46 (2008).
26. Vandekerchove, D., P.D. Herdt, H. Laevens and F. Pasmans, Risk factors associated with colibacillosis outbreaks in caged layer flocks. *Avian Pathol.*, **33**: 337-342 (2004 a).
27. Vandekerchove, D., P. DeHerdt, H. Laevens, P. Butaye, G. Meulemans and F. Pasmans., Significance of interactions between *Escherichia coli* and respiratory pathogens in layer hen flocks suffering from colibacillosis- associated mortality. *Avian Pathol.*, **33**: 298-302 (2004 b).
28. Vijayalingam, T.A., G.A. Balasubramanian, C. Balachandran and V. Titus George, Pathology of ovary and oviduct in spontaneous cases of colibacillosis in layer chicken. *Indian J. Vet. Pathol.*, **31**: 178-179 (2007).
29. Zhao, C., B.Ge, J.D. Villena, R. Sudler, E. Yeh, S. Zhao, D.G. White, D. Wagner and J. Meng, Prevalence of *Campylobacter spp.*, *Escherichia coli* and *Salmonella* serovars in retail chicken, turkey, pork and beef from the Great Washington, D.C., Area. *App. Environ. Microbiol.*, **67**: 5431-5436 (2001).